

Supplemental Figure. 1. Effect of medium conditioning by ML265-treated cells on T cell activation. Macrophages from CAD patients were stimulated with LPS/IFN- γ in the absence or presence of ML265. After 12 hrs, culture medium was replaced by fresh medium. After 24 hrs, culture medium was transferred to naïve CD4 T cells, which were activated with CD3/CD28-coated beads at a ratio of 1:4. After 48 hrs, the surface expression of CD25 and CD69 was measured by flow cytometry. (A) Representative flow cytometry dot plots. (B) Summary of 6 experiments. The data are shown as mean \pm SEM. n.s.; not significant. Paired 2tailed Student's *t* test was used for comparisons.



Supplemental Figure 2. Influence of medication exposure and gender on PD-L1 expression. (A, B) Macrophages were generated from individuals without CAD treated with triple therapy (aspirin, a statin, and an angiotensin-converting enzyme inhibitor). Individuals on no medication served as controls. Macrophages were stimulated with LPS/IFN-γ and surface expression of PD-L1 was measured by flow cytometry after 24 hrs. (A) Representative histograms. (B) Mean fluorescence intensity (MFI) from n=10 experiments. (C) Macrophages were generated from 25 male CAD patients and 9 female CAD patients. Macrophages were stimulated with LPS/IFN-γ and surface expression of PD-L1 was measured by flow cytometry after 24 hrs. All data are mean ± SEM. n.s.; not significant. The Mann-Whitney test was used for comparisons.



Supplemental Figure 3. siRNA-knockdown of PD-L1. Macrophages from CAD patients transfected with either siControl or siPD-L1 RNA for 48 hrs were stimulated with LPS/IFN- γ for 24 hrs. The surface expression of PD-L1 and PD-L2 was measured by flow cytometry. Representative histograms are shown.



Supplemental Figure 4. BMP4-IRF1-PD-L1 Upregulation in CAD Monocytes. CD14⁺ monocytes from healthy individuals and CAD patients were isolated from freshly harvested peripheral blood. **(A)** Gene expressions of BMP4 were measured by RT-PCR. **(B)** Gene expressions of IRF1 were measured by RT-PCR. **(C, D)** Surface expression of PD-L1 was analyzed by flow cytometry. **(C)** Representative histograms of PD-L1 expression. **(D)** Mean \pm SEM of the PD-L1 MFI from 6 independent experiments. **P* < 0.05; ***P* < 0.01. The Mann-Whitney test was used for comparisons.



Supplemental Figure 5. Recombinant BMP4 induces IRF1 and PD-L1. Macrophages from healthy individuals were stimulated with LPS/IFN- γ in the absence or presence of recombinant BMP4 (10 ng/ml) for 24 hrs. Intracellular IRF1 and surface expression of PD-L1 analyzed by flow cytometry. Data are from 6 experiments. All data are mean ± SEM. **P* < 0.05; ***P* < 0.01. Paired 2-tailed Student's *t* test was used for comparisons.

Genes	Forward primers	Reverse primers
PD-L1	GGTTGTGGATCCAGTCACCT	TTGGTGGTGGTGGTCTTACC
PD-L2	CAGTGCTATCTGAACCTGTGGT	CTGCAGGCCACCGAATTCTT
CD80	ATGGTGGGCACAGAAGTAGC	AGGAAATCTGGGTTCTGGCG
CD86	TGGTCAGGGAGGGGTTTTGG	GCCCCGGGTGATCTGTGTCT
ICOSL	GCAGCCTTCGAGCTGATACTC	GTTTTCGACTCACTGGTTTGC
OX40L	CACCTACATCTGCCTGCACTTC	TTCGGTAAATTGTACTTTGATACT
BMP4	ATGATTCCTGGTAACCGAATGC	CCCCGTCTCAGGTATCAAACT
IRF1	CTGTGCGAGTGTACCGGATG	ATCCCCACATGACTTCCTCTT
β-actin	GATCATTGCTCCTCCTGAGC	CGTCATACTCCTGCTTGCTG

Suppl. Table 1. Primers used in the manuscript